

# RISE-Meta: Meta-analytic Evaluation of High-Dimensional Trial-Level Surrogates Applied to Vaccinology

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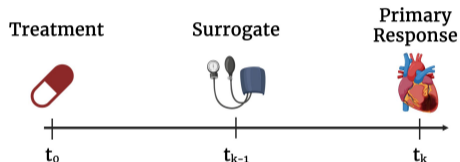
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## Surrogate Markers

# What is a surrogate marker?

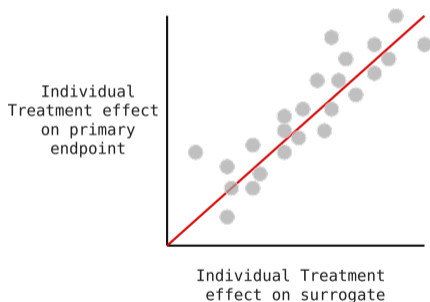
- **Intermediate endpoint**
- Treatment effect on surrogate **predicts treatment effect(s) on primary endpoint**



Treatment	Primary endpoint	Surrogate	Advantage
Antiretroviral therapy	Progression to AIDS	Plasma HIV-1 viral load	Observed sooner
Statin therapy	Major cardiovascular events	Reduction in LDL cholesterol	Easy to measure, observed sooner
Yellow fever vaccination	Yellow fever infection	Neutralising antibody titre	Measuring vaccine response when disease is not endemic

# Individual versus trial-level surrogates

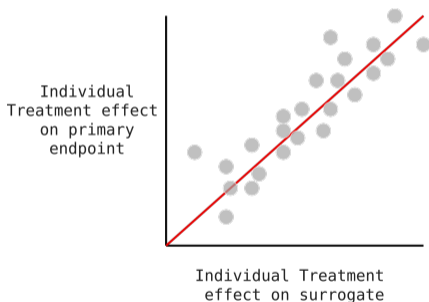
## Individual-Level Surrogate



- Evaluated with correlation, association, cross-validation... (Gabriel, 2017)

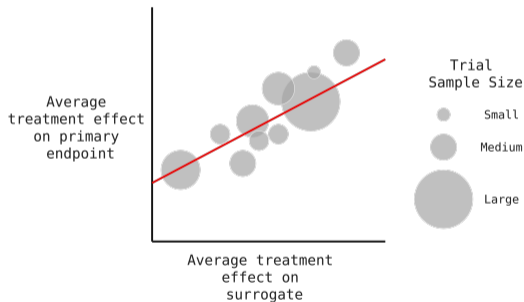
# Individual versus trial-level surrogates

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## Trial-Level Surrogate

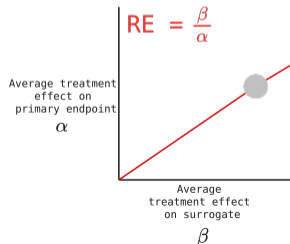


- Evaluated in *single-trial* or *multi-trial* framework

# Evaluation of Trial-Level Surrogacy

## Single-Trial Framework

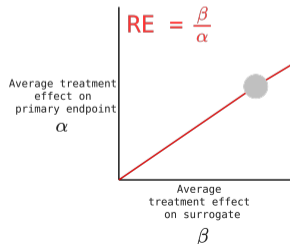
- Proportion of treatment effect explained (Freedman, 1992), relative effect (Buyse, 2000), nonparametric approaches (Parast, 2024; Hughes, 2025)
- **Implicit assumption about portability**



# Evaluation of Trial-Level Surrogacy

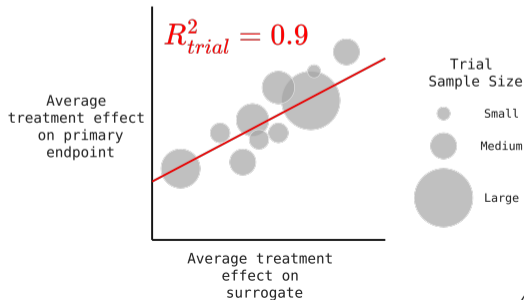
## Single-Trial Framework

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- **Implicit assumption about portability**



## Multi-Trial Framework

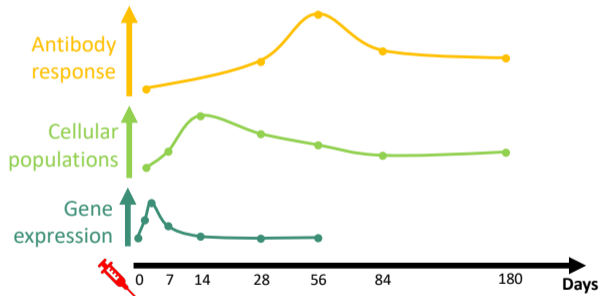
- Targets **generalisability** of surrogate across clinical settings
- Evaluated with meta-analysis (Molenberghs, 2002)



## High-Dimensional Surrogates

# High-dimensional data in biomedical research

- High-throughput technologies: **thousands of variables per sample**
  - e.g. transcriptomics, proteomics, metabolomics
- Potential as **biomarkers** of downstream endpoints
- Literature focused on differential analysis and individual prediction, **surrogates previously unexplored**



Credit: Boris Hejblum

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## Previously existing methods

- Restrictive assumptions
- Large sample sizes
- Low-dimensional

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## Rank-based Identification of high-dimensional SurrogatE markers (RISE) (Hughes, 2025)

- Nonparametric evaluation of high-dimensional surrogates in small sample setting
- Single-trial framework

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## Rank-based Identification of high-dimensional SurrogatE markers (RISE) (Hughes, 2025)

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- Single-trial framework

**Goal:** Extend RISE to meta-analytic setting  
Identify **generalisable high-dimensional surrogates**

RISE

# Notation

## Data structure

- $m = 1, \dots, M$  : trials
- $i = 1, \dots, N_m$  : individuals in trial  $m$
- $j = 1, \dots, J$  : candidate surrogate markers

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## Variables

- Binary treatment  $A_{i,m} \in \{0, 1\}$
- Primary endpoint  $Y_{i,m}^a$
- Surrogate vector  $\mathbf{S}_{i,m}^a = (S_{i,1,m}^a, \dots, S_{i,J,m}^a)$

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## Observed data

- $(A_{i,m}, Y_{i,m}^{A_{i,m}}, \mathbf{S}_{i,m}^{A_{i,m}}) \forall i$
- independent, i.i.d. within trial  $m$

# Within-trial effect measures

## Treatment effects with U-statistics (Parast, 2024; Hughes, 2025)

$$U_{Y,m} = \mathbb{P}(Y_{i,m}^1 > Y_{l,m}^0) + \frac{1}{2}\mathbb{P}(Y_{i,m}^1 = Y_{l,m}^0)$$

$$U_{S,m} = \mathbb{P}(S_{i,m}^1 > S_{l,m}^0) + \frac{1}{2}\mathbb{P}(S_{i,m}^1 = S_{l,m}^0)$$

- **Probability** that treated individual has *greater* response than control
  - $\Rightarrow > 0.5$ : positive effect     $= 0.5$ : no effect     $< 0.5$ : negative effect

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### Within-trial surrogacy

- $\delta_m = U_{Y,m} - U_{S,m}$ 
  - **bias in estimating effect on  $Y$  observing only  $S$**
- **Quantity of interest for meta-analysis!**

# Within-Trial Estimation

$$G(x, y) = \begin{cases} 1, & x > y \\ \frac{1}{2}, & x = y \\ 0, & x < y \end{cases}$$

## Active treatment vs control

$$\hat{U}_{Y,m} = \frac{1}{N_m^1 N_m^0} \sum_{i=1}^{N_m^1} \sum_{l=1}^{N_m^0} G(Y_{i,m}^1, Y_{l,m}^0), \quad \hat{U}_{S,m} = \frac{1}{N_m^1 N_m^0} \sum_{i=1}^{N_m^1} \sum_{l=1}^{N_m^0} G(S_{i,m}^1, S_{l,m}^0)$$

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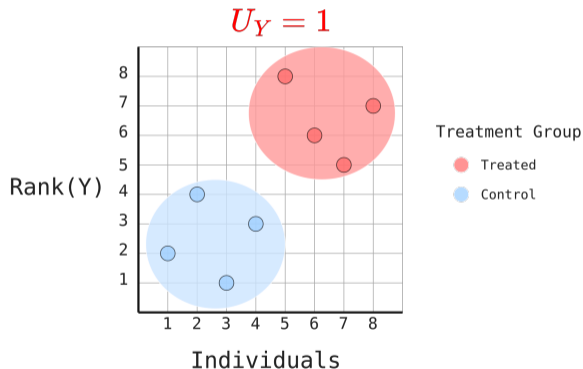
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## Paired setting

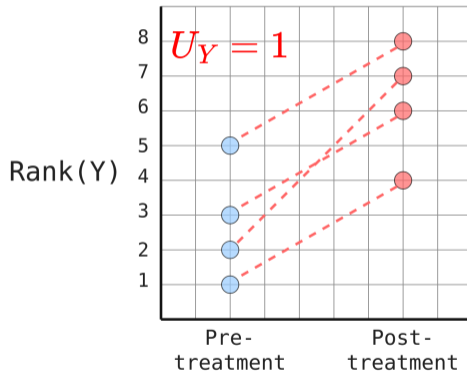
$$\hat{U}_{Y,m} = \frac{1}{N_m} \sum_{i=1}^{N_m} G(Y_{i,m}^1, Y_{i,m}^0), \quad \hat{U}_{S,m} = \frac{1}{N_m} \sum_{i=1}^{N_m} G(S_{i,m}^1, S_{i,m}^0)$$

# Intuition for estimation - Active versus control



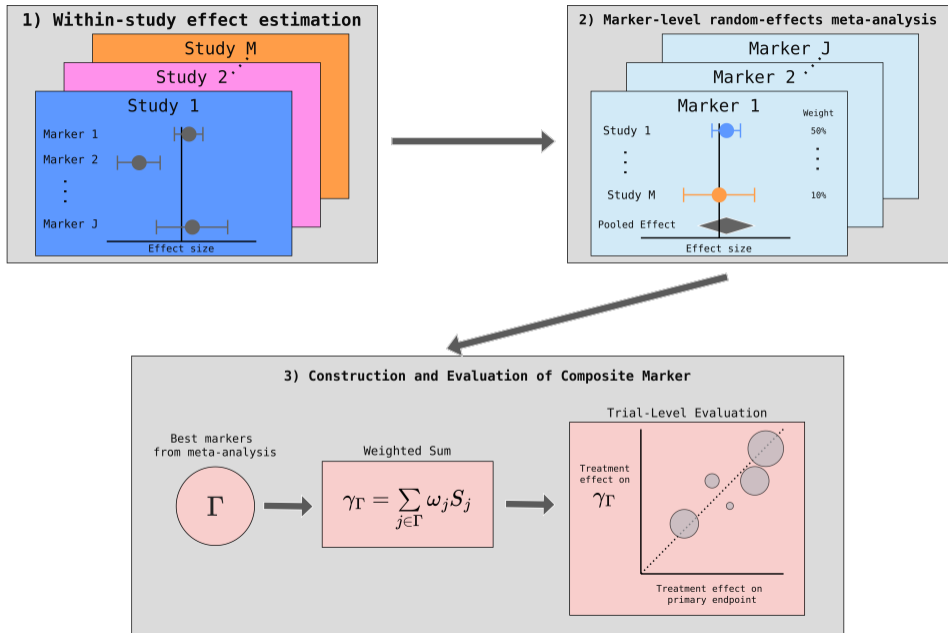
$\hat{U}_{Y,m} = 1 \implies$  **perfect group separation by  $Y$  in trial  $m$**

# Intuition for estimation - paired case



$\hat{U}_{Y,m} = 1 \implies$  response improves with treatment for every individual in trial  $m$

RISE-Meta



# Random effects meta-analysis model

Assume within-trial effects  $\delta_m = U_{Y,m} - U_{S,m}$  are gaussian (DerSimonian, 1986)

$$\delta_m \sim N(\mu_\delta, \sigma_{\delta,m}^2 + \tau_\delta^2)$$

where

- $\mu_\delta$ : common mean effect
- $\sigma_{\delta,m}^2$ : study-specific variance in trial  $m$
- $\tau_\delta^2$ : common between-study variance
  - Estimated with REML

# Estimating the mean effect

Estimate **pooled effect** as **weighted sum of within-trial estimates**

$$\widehat{\mu}_{\delta} = \frac{\sum_{m=1}^M \widehat{w}_{\delta,m} \widehat{\delta}_m}{\sum_{m=1}^M \widehat{w}_{\delta,m}}$$

**weights** are **inverse variances**

$$\widehat{w}_{\delta,m} = \frac{1}{\widehat{\sigma}_{\delta,m}^2 + \widehat{\tau}_{\delta}^2}$$

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- Recommend variance estimated with Hartung-Knapp estimator  $\widehat{\text{Var}}_{\text{HK}}$  (Hartung, 2001)
- $(1 - \alpha) \times 100\%$  C.I. :  $\widehat{\mu}_{\delta} \pm q_{t_{M-1}, 1-\alpha/2} \sqrt{\widehat{\text{Var}}_{\text{HK}}(\widehat{\mu}_{\delta})}$

# Hypothesis testing on pooled effect

**Goal: test surrogacy with**  $\mu_\delta \in [-\varepsilon, \varepsilon]$

Propose a *two one-sided test* (TOST) procedure to test (Schuirmann, 1987)

$$H_{0L}^{\text{TOST}} : \mu_\delta \leq -\varepsilon, \quad H_{0U}^{\text{TOST}} : \mu_\delta \geq \varepsilon$$

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$$H_{0L}^{\text{TOST}} : \mu_\delta \leq -\varepsilon, \quad H_{0U}^{\text{TOST}} : \mu_\delta \geq \varepsilon$$

⇒ two sets of test statistics and p-values

$$T_L = \frac{\hat{\mu}_\delta + \varepsilon}{\widehat{\text{SE}}_{HK}}, \quad p_L = 1 - F_{t, M-1}(T_L),$$

$$T_U = \frac{\hat{\mu}_\delta - \varepsilon}{\widehat{\text{SE}}_{HK}}, \quad p_U = 1 - F_{t, M-1}(T_U),$$

$F_{t, M-1}$  is c.d.f. of t-distribution with  $M - 1$  degrees of freedom. Combined p-value is

$$p_{\text{TOST}} = \max(p_L, p_U)$$

# Deriving a signature of promising markers

- $p_j$  = p-value for  $j$ th marker testing  $\hat{\mu}_{\delta_j} \in [-\varepsilon, \varepsilon]$
- Apply correction to control for false discovery rate (Benjamini, 1995)
- $p_{adj,j}$  = adjusted p-value for  $j$ th marker

Define a signature set

$$\Gamma = \{j : p_{adj,j} \leq \alpha\}$$

# Constructing a composite surrogate signature

**Goal:** combine markers  $S_j : j \in \Gamma$  into a univariate signature

- **Surrogacy strength component:**  $\hat{a}_j = \frac{\varepsilon - |\hat{\mu}_{\delta,j}|}{\varepsilon}$

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**Composite signature:**

$$\hat{\gamma}_{\Gamma} = \sum_{j \in \Gamma} \hat{\omega}_j \bar{S}_j$$

# Prediction intervals

- **Confidence intervals misleading** in presence of between-trial **heterogeneity** (IntHout, 2016)
  - Good trial-level surrogate must have  $\mu_\delta \approx 0$  **and consistent estimation** across trials

# Prediction intervals

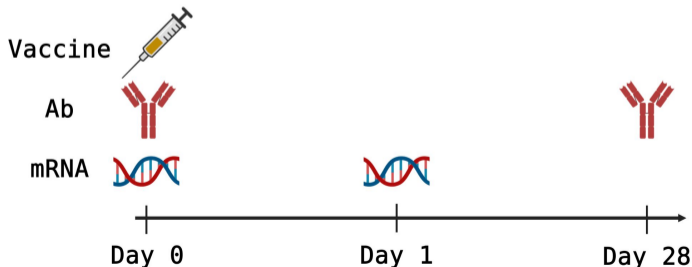
- **Confidence intervals misleading** in presence of between-trial **heterogeneity** (IntHout, 2016)
  - Good trial-level surrogate must have  $\mu_\delta \approx 0$  **and consistent estimation** across trials
- **Prediction interval**: likely range of effect estimates in **new trials**
- $(1 - \alpha) \times 100\%$  P.I. given by

$$\widehat{\mu}_\delta \pm q_{t_{M-1}, 1-\alpha/2} \sqrt{\widehat{\text{Var}}_{HK}(\widehat{\mu}_\delta) + \widehat{\tau}_\delta^2}$$

Application

# Application to inactivated influenza vaccine

- **Open data** from **4 trials** in Human Immune Project Consortium on inactivated flu vaccine (Diray-Arce, 2022)
- **Primary endpoint:** Day 28 antibody response (nAb/HAI)
- **Surrogate candidates** : gene expression of  $\approx 10,000$  genes
- **Paired setting** (each individual pre- & post-vaccination Ab and GE)



Can we use RISE-Meta to derive a **generalisable signature** of **gene expression**, measured at 1 day after vaccination, which is **predictive of the vaccine effect** on the **antibodies**, measured at 28 days post-vaccination?

# Analysis Details

- **Transform data gene-level  $\Rightarrow$  geneset-level**
  - Transformation = within-geneset mean expression
  - $\Rightarrow$  **reduce dimension**, improve interpretability
  - Genesets: blood transcriptional modules ( $J = 174$  after removing untitled) (Li, 2013)

# Analysis Details

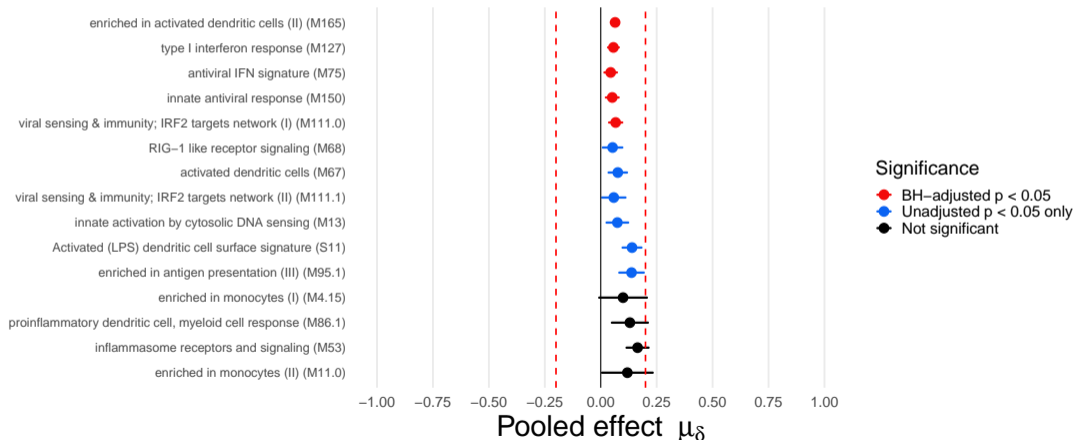
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  - Marker-level meta-analyses on training data ( $\frac{2}{3}$  within each trial)
  - Evaluate composite marker on test data

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  - Marker-level meta-analyses on training data ( $\frac{2}{3}$  within each trial)
  - Evaluate composite marker on test data
- Hyperparameters :  $\varepsilon = 0.2$ ,  $\alpha = 0.05$ , Benjamini-Hochberg correction

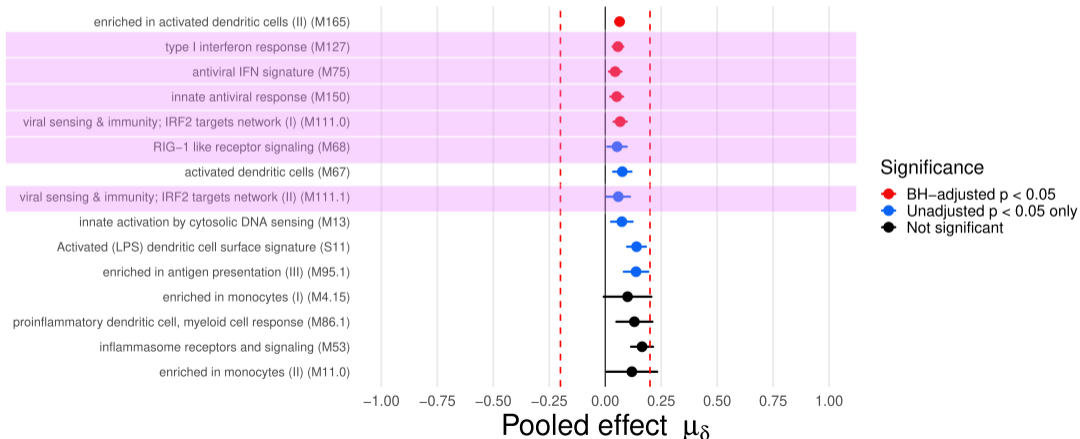
# Interferon and dendritic cell signals dominate the signature

## Screening results: Top 15 markers by p-value



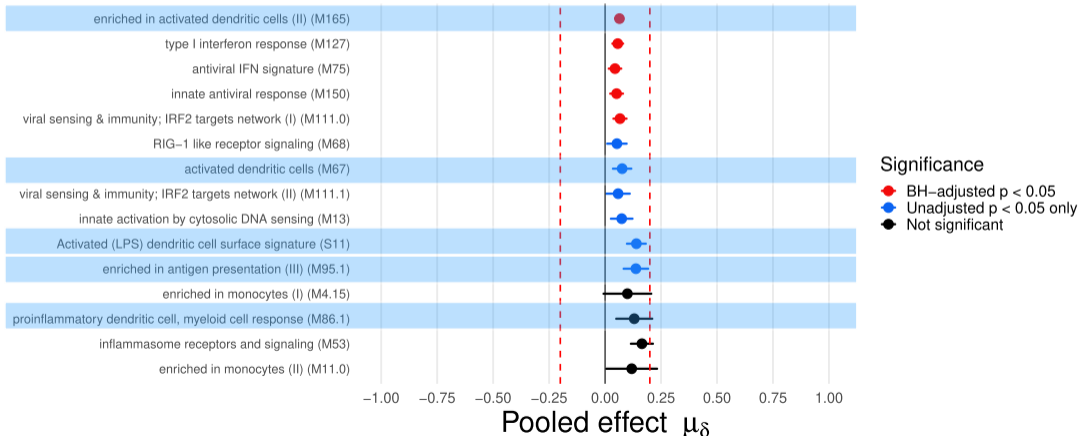
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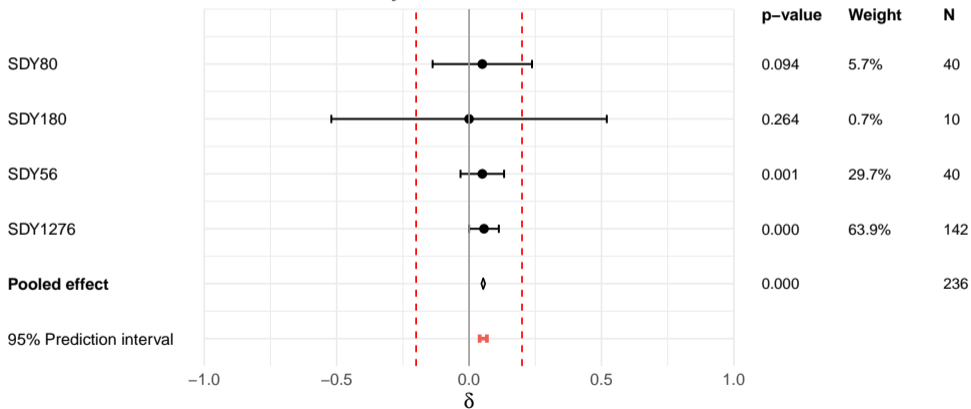
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# Evaluation of composite signature on independent data

Random-effects meta-analysis of combined marker in evaluation data



Tau-squared = 0.0000 | I-Squared = 0.0% | Lin's CCC = 0.85 | k = 4

## Discussion

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- RISE: statistical framework for high-dimensional surrogate evaluation in a **single-trial**
- **Meta-analysis: gold-standard** for evaluation of surrogacy considering **between-study heterogeneity**
- Application: **promising signature** of biologically relevant genes **predictive of flu vaccine effect** on the later antibody response

**RISE-Meta** is a novel statistical framework for the **meta-analysis of high-dimensional trial-level surrogate markers**

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## Perspectives

- ⇒ Extension to other outcome types (binary, survival)
- ⇒ Simulations to explore limits (number of studies, study size)
- ⇒ Bayesian meta-analysis when number of trials is small?

# Thank you!

RISE Available in the **R package** SurrogateRank








RISE-Meta available soon







Hughes A, Parast L, Thiébaud R & Hejblum BP. RISE: Two-stage rank-based identification of high-dimensional surrogate markers applied to vaccinology. *Statistics in Medicine* 44(20-22):e70241, 2025.  
DOI: 10.1002/sim.70241





# References I

-  Ball, Tali M. (Mar. 2020). “Double Dipping in Machine Learning: Problems and Solutions”. In: *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging* 5.3, pp. 261–263. ISSN: 2451-9022.
-  Benjamini, Yoav and Yosef Hochberg (Jan. 1995). “Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing”. In: *Journal of the Royal Statistical Society Series B: Statistical Methodology* 57.1, pp. 289–300. ISSN: 1467-9868.
-  Buyse, Marc (Apr. 2000). “Statistical Validation of Surrogate Endpoints: Problems and Proposals”. In: *Drug Information Journal* 34.2, pp. 447–454. ISSN: 2164-9200.
-  DerSimonian, Rebecca and Nan Laird (Sept. 1986). “Meta-analysis in clinical trials”. In: *Controlled Clinical Trials* 7.3, pp. 177–188. ISSN: 0197-2456.
-  Diray-Arce, Joann (Oct. 2022). “The Immune Signatures data resource, a compendium of systems vaccinology datasets”. In: *Scientific Data* 9.1. ISSN: 2052-4463.

# References II

-  Freedman, Laurence S., Barry I. Graubard, and Arthur Schatzkin (Jan. 1992). “Statistical validation of intermediate endpoints for chronic diseases”. In: *Statistics in Medicine* 11.2, pp. 167–178. ISSN: 1097-0258.
-  Gabriel, Erin E, Michael C Sachs, and M Elizabeth Halloran (Sept. 2017). “Evaluation and comparison of predictive individual-level general surrogates”. In: *Biostatistics* 19.3, pp. 307–324. ISSN: 1468-4357.
-  Hartung, Joachim and Guido Knapp (Dec. 2001). “A refined method for the meta-analysis of controlled clinical trials with binary outcome”. In: *Statistics in Medicine* 20.24, pp. 3875–3889. ISSN: 1097-0258.
-  Hughes, Arthur (Sept. 2025). “RISE: Two-Stage Rank-Based Identification of High-Dimensional Surrogate Markers Applied to Vaccinology”. In: *Statistics in Medicine* 44.20–22. ISSN: 1097-0258.

## References III

-  IntHout, Joanna (July 2016). “Plea for routinely presenting prediction intervals in meta-analysis”. In: *BMJ Open* 6.7, e010247. ISSN: 2044-6055.
-  Li, Shuzhao (Dec. 2013). “Molecular signatures of antibody responses derived from a systems biology study of five human vaccines”. In: *Nature Immunology* 15.2, pp. 195–204. ISSN: 1529-2916.
-  Molenberghs, Geert (Dec. 2002). “Statistical challenges in the evaluation of surrogate endpoints in randomized trials”. In: *Controlled Clinical Trials* 23.6, pp. 607–625. ISSN: 0197-2456.
-  Parast, Layla, Tianxi Cai, and Lu Tian (Jan. 2024). “A rank-based approach to evaluate a surrogate marker in a small sample setting”. In: *Biometrics* 80.1. ISSN: 1541-0420.

## References IV



Schuirmann, Donald J. (Dec. 1987). “A comparison of the Two One-Sided Tests Procedure and the Power Approach for assessing the equivalence of average bioavailability”. In: *Journal of Pharmacokinetics and Biopharmaceutics* 15.6, pp. 657–680. ISSN: 0090-466X.

## Data Description

# Available studies

Study	Year	Study Description
SDY61	2008	Systems Biology of 2007 Influenza Vaccination in Humans
SDY269	2009	Systems Biology of 2008 Influenza Vaccination in Humans
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SDY180	2010	Systems scale interactive exploration reveals quantitative and qualitative differences in response to 2009-2010 Fluzone influenza vaccine and pneumococcal vaccine
SDY56	2010	Systems Biology of 2010 trivalent Influenza vaccine (TIV) in young and elderly
SDY67	2010	Bioinformatics Approach to 2010-2011 TIV Influenza A/H1N1 Vaccine Immune Profiling
SDY1119	2011	Systems Biology of 2011 trivalent Influenza vaccine (TIV) in young and elderly individuals, healthy or with T2D
SDY224	2011	Immune Responses to Seasonal TIV 2010-2011 Influenza Vaccination in Humans
SDY404	2011	Immunologic and genomic signatures of influenza vaccine response - 2011
SDY63	2011	Immunologic and genomic signatures of influenza vaccine response - 2010
SDY400	2012	Immunologic and genomic signatures of influenza vaccine response - 2012
SDY1276	2013	Time series of global gene expression after trivalent influenza vaccination in humans
SDY520	2013	Immunologic and genomic signatures of influenza vaccine response - 2013
SDY80	2014	Cellular and molecular characterization of the immune response in healthy NIH employees at baseline, and after immunization with the H1N1 or seasonal influenza vaccines
SDY640	2014	Immunologic and genomic signatures of influenza vaccine response - 2014

# Inclusion/Exclusion Criteria

Study	Inclusion	Exclusion
SDY61	Healthy adults	
SDY269	Healthy adults	
SDY270	Healthy adults	
SDY180	Healthy adults (18-64)	Pregnancy, allergies, vaccines within 2 months
SDY56	Healthy adults (18-40 or $\geq 65$ )	Recent infections/vaccines, chronic illnesses, pregnancy
SDY67	Healthy adults (50-74)	
SDY1119	Healthy or <b>type 2 diabetic</b> adults, immunocompetent	Other illnesses, pregnancy, drug intake
SDY224	Healthy adults (18-69)	Other illnesses, pregnancy, drug intake
SDY404	Healthy adults (21-30 or $\geq 65$ )	
SDY63	Healthy adults (21-30 or $\geq 65$ )	
SDY400	Healthy adults (21-30 or $\geq 65$ )	
SDY1276	Healthy adults (18-40), white	Influenza infection or vaccine within 3 years
SDY520	Healthy adults (21-30 or $\geq 65$ )	
SDY80	Healthy adults, NIH employee	
SDY640	Healthy adults (21-30 or $\geq 65$ )	

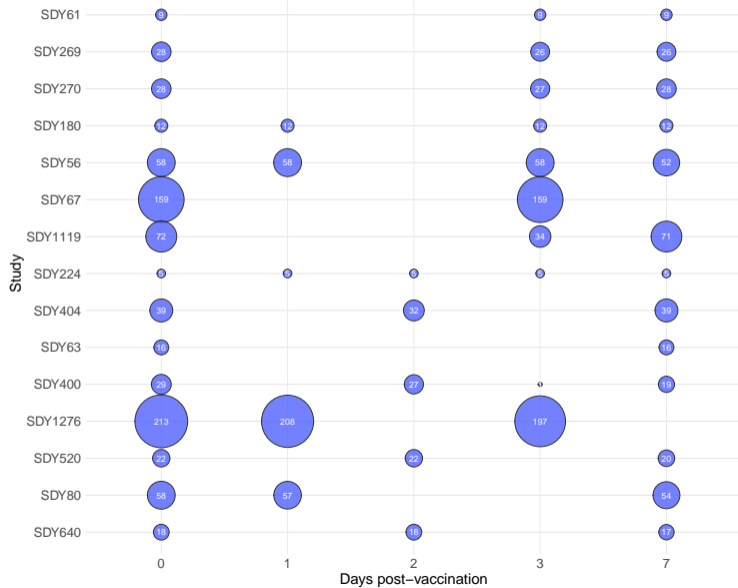
# Inclusion criteria for surrogate analysis

Participants must:

- be healthy adults (18+),
- receive inactivated influenza vaccine,
- have at least one immune response assay measurement (nAb, HAI) at baseline AND day 28 (+/-7 days)
- have gene expression measured both at baseline AND post-vaccination

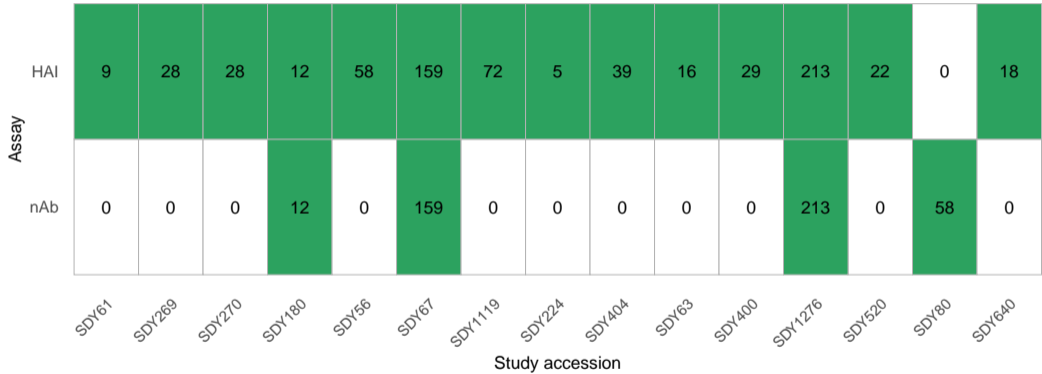
## Number of transcriptomic observations across time per study

Selected timepoints, healthy adults with baseline and post-vaccination data



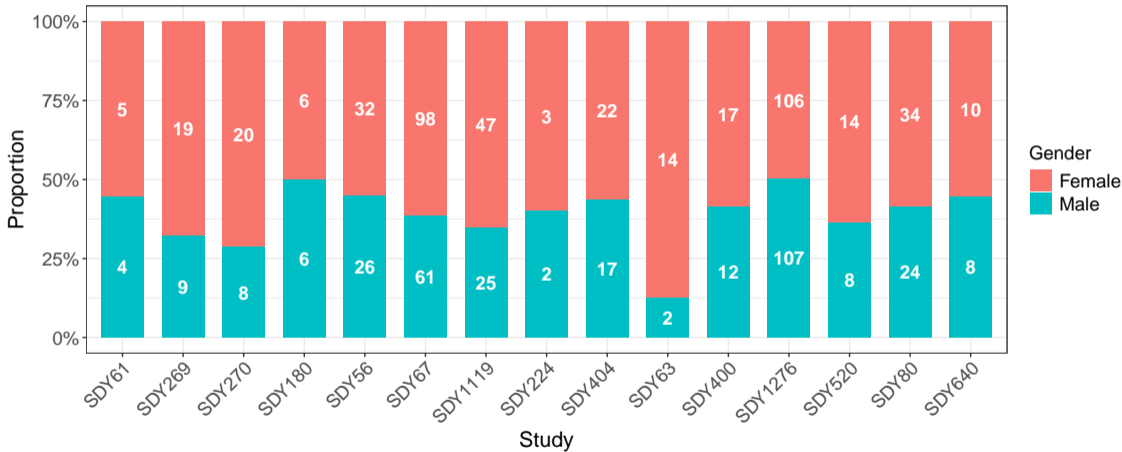
## Availability of antibody assays per study

Participants with both baseline and day 28 (+/- 7 days) post-vaccination measurements

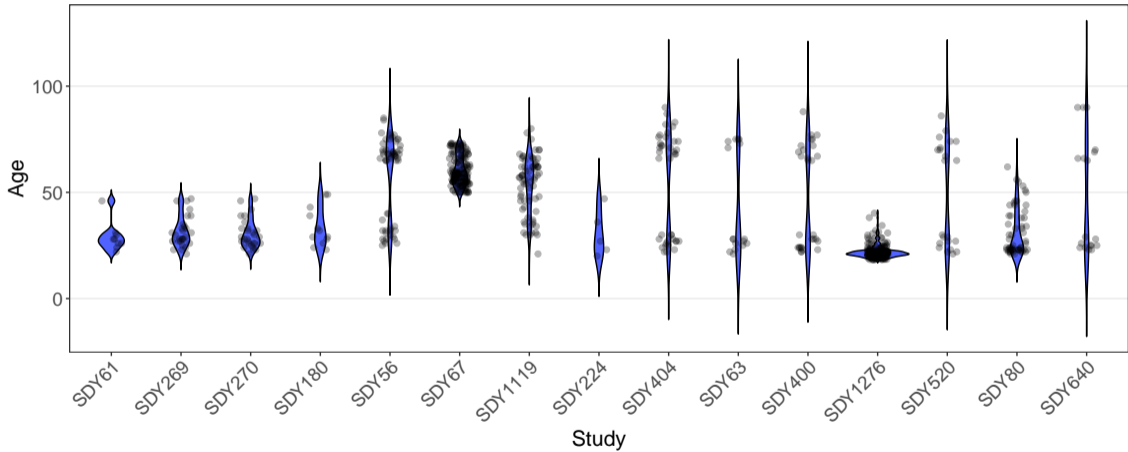


- Use nAb if available, otherwise HAI

### Gender distribution by study



## Age distribution by study



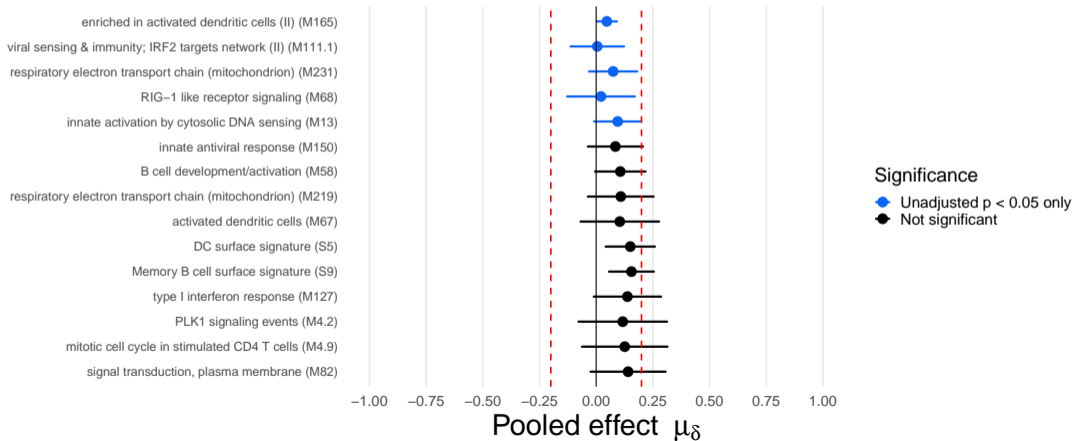
## Supplementary Analyses

# Supplementary analyses

- Day 1, 2, 3, 7 post-vaccination
- Genewise analyses

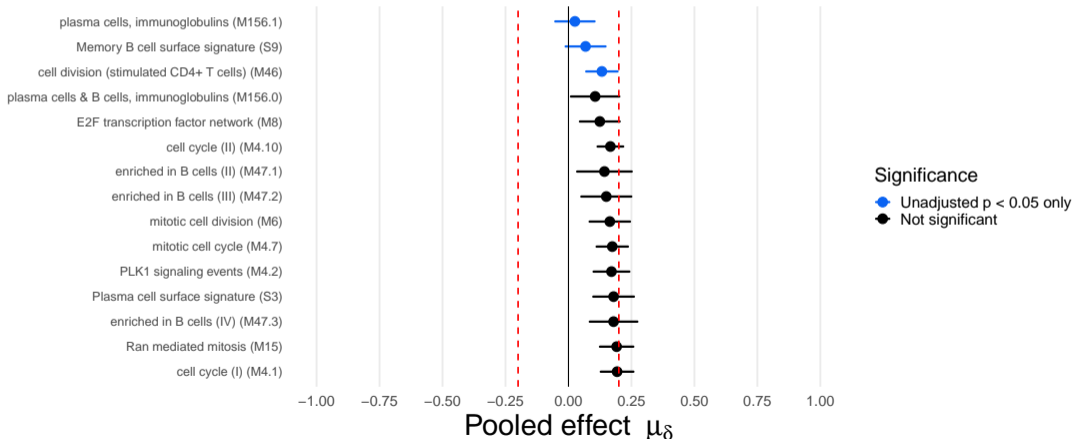
# Day 2 screening results

## Screening results: Top 15 markers by p-value



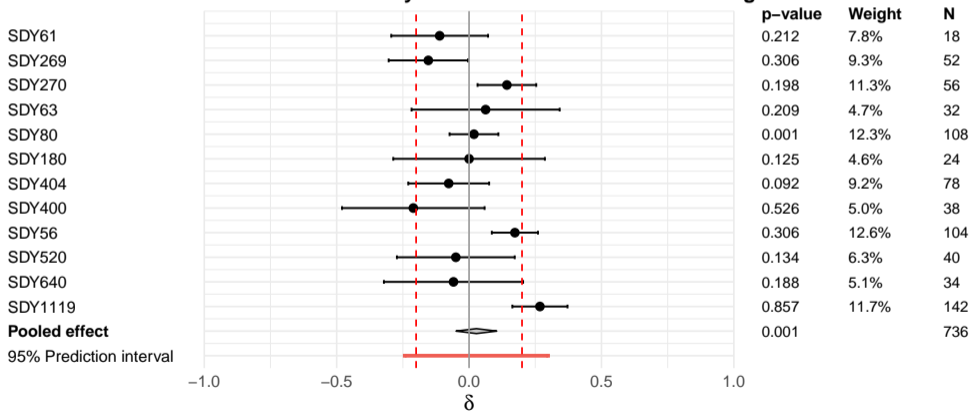
# Day 7 screening results

## Screening results: Top 15 markers by p-value



# Day 7 evaluation results

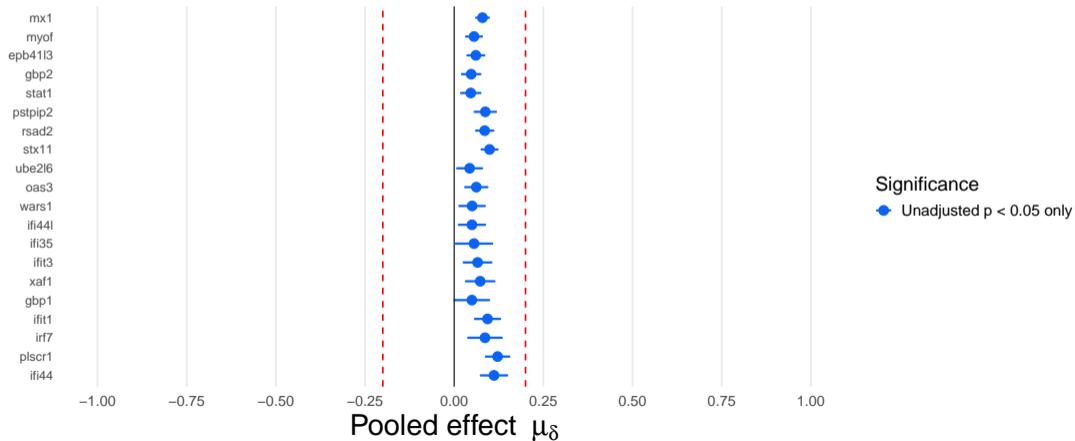
## Random-effects meta-analysis of combined marker in training data



Tau-squared = 0.0129 | I-Squared = 64.8% | Lin's CCC = 0.21 | k = 12

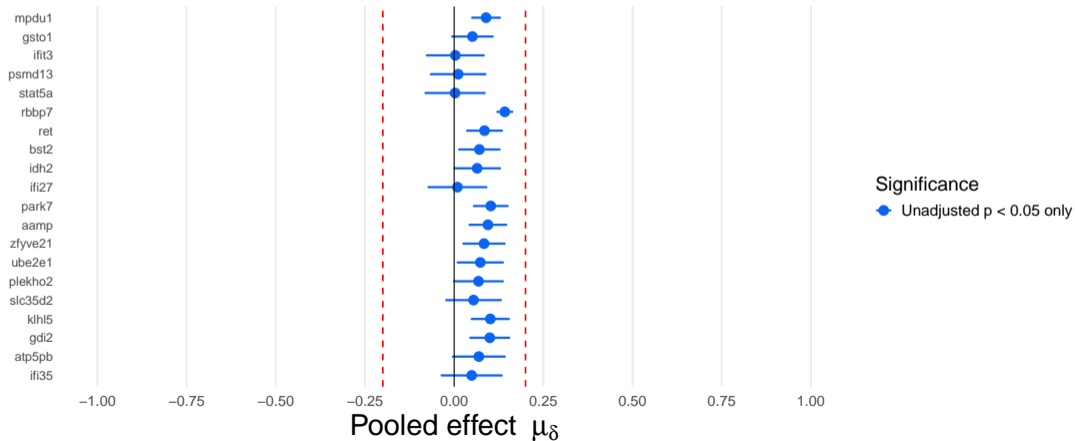
# Day 1 Genewise Analysis

## Screening results: Top 20 markers by p-value



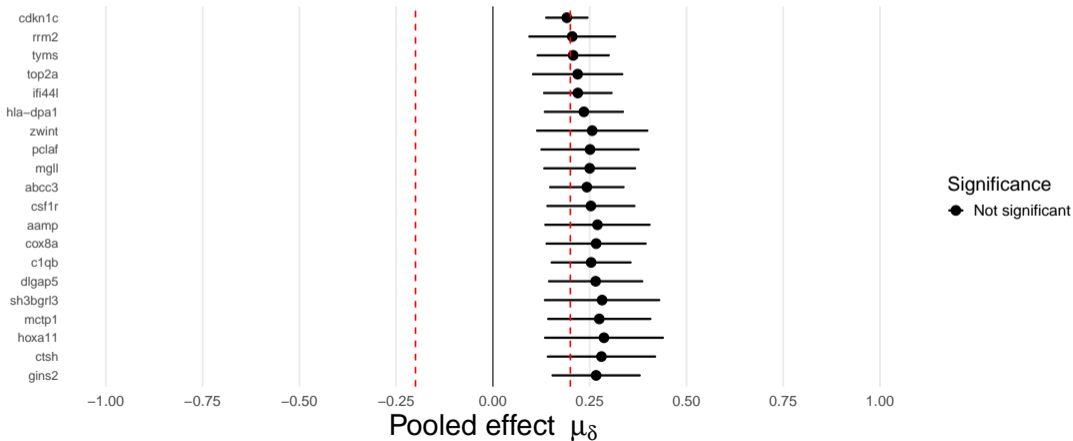
# Day 2 Genewise Analysis

## Screening results: Top 20 markers by p-value



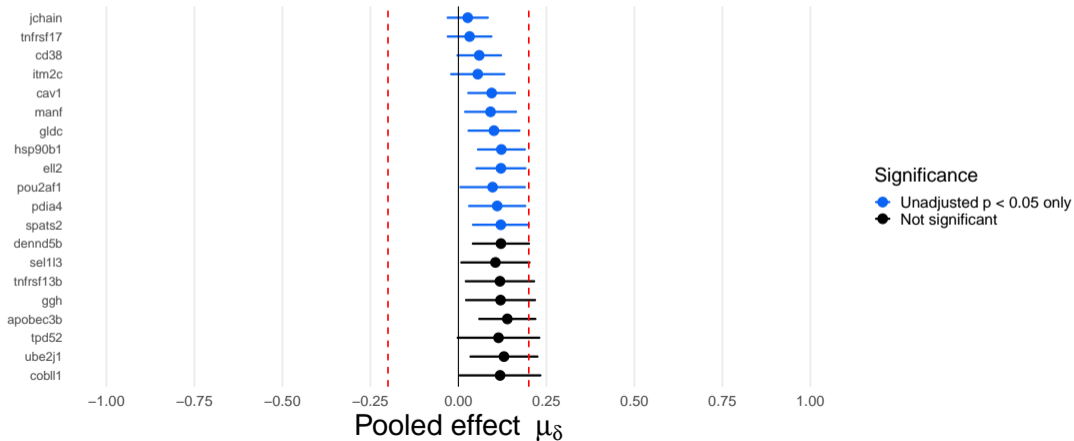
# Day 3 Genewise Analysis

## Screening results: Top 20 markers by p-value



# Day 7 Genewise Analysis

## Screening results: Top 20 markers by p-value



Simulation

# Gaussian simulation

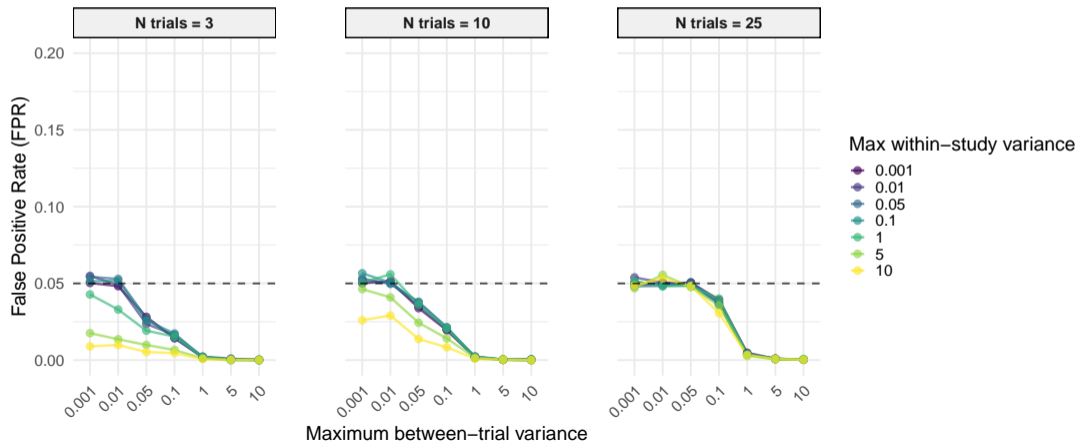
- Simulate true deltas:  $\delta_{m,j}^{\text{true}} \sim \mathcal{N}(\mu_{\delta,j}, \tau_{\delta,j}^2)$ 
  - allow heterogeneity to vary across markers :  
 $\tau_{\delta,j}^2 \sim \mathcal{U}(u_{\tau,\min}, u_{\tau,\max}), \quad 0 \leq u_{\tau,\min} \leq u_{\tau,\max}$
- add sampling error :  $\hat{\delta}_{m,j} \sim \mathcal{N}(\delta_{m,j}^{\text{true}}, \sigma_{m,j}^2)$ 
  - where  $\sigma_{m,j}^2 = \frac{\nu_j}{n_m}$  has trial-specific component  $n_m > 0$  and marker-specific component  
 $\nu_j \sim \mathcal{U}(u_{\nu,\min}, u_{\nu,\max}), \quad 0 < u_{\nu,\min} \leq u_{\nu,\max}$

# Definition of surrogate validity

- $S_j$  valid if  $\mu_{\delta,j} \in (-\varepsilon, \varepsilon)$
- $S_j$  invalid if :  $\mu_{\delta,j} \in [-1, -\varepsilon] \cup [\varepsilon, 1]$
- To examine calibration, generate invalid surrogates in "worst-case" (i.e. on boundary defined by  $\varepsilon$ )
  - Better case scenarios will be at least as well calibrated

# Calibration

## Calibration – false positive rate across different settings



# Power

## Power – true positive rate across different settings

